Skeletal muscle changes after hemiparetic stroke and potential beneficial effects of exercise intervention strategies

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Abstract—Stroke is the leading cause of disability in the United States. New evidence reveals significant structural and metabolic changes in skeletal muscle after stroke. Muscle alterations include gross atrophy and shift to fast myosin heavy chain in the hemiparetic (contralateral) leg muscle; both are related to gait deficit severity. The underlying molecular mechanisms of this atrophy and muscle phenotype shift are not known. Inflammatory markers are also present in contralateral leg muscle after stroke. Individuals with stroke have a high prevalence of insulin resistance and diabetes. Skeletal muscle is a major site for insulin-glucose metabolism. Increasing evidence suggests that inflammatory pathway activation and oxidative injury could lead to wasting, altered function, and impaired insulin action in skeletal muscle. The health benefits of exercise in disabled populations have now been recognized. Aerobic exercise improves fitness, strength, and ambulatory performance in subjects with chronic stroke. Therapeutic exercise may modify or reverse skeletal muscle abnormalities.

Key words: body composition, exercise, inflammation, insulin-glucose metabolism, myosin heavy chain isoforms, rehabilitation, sarcopenia, skeletal muscle, stroke, walking.

INTRODUCTION

Stroke is the leading cause of chronic disability in the United States, and hemiparesis is the most common chronic disabling sequela after stroke [1–3]. A paucity of literature exists on skeletal muscle abnormalities and their clinical relevance after stroke. The majority of conventional stroke rehabilitation occurs during the subacute period. Unfortunately, little rehabilitation care is prescribed during the chronic stroke phase. Few or no evidence-based recommendations promote regular exercise after stroke. Moreover, the conventional physical therapy administered during the subacute recovery period likely does not provide adequate exercise stimulus to reverse the potential skeletal muscle abnormalities or deconditioning that follow stroke [4–5]. Skeletal muscle has not been systematically pursued as a potential target for exercise and/or rehabilitation after stroke. This article outlines our current knowledge on (1) alterations of body composition and muscle structure and function in aging, inactivity, and after spinal cord injury (SCI); (2) alterations of muscle structure and function

Abbreviations: ATPase = adenosine triphosphatase, DXA = dual X-ray absorptiometry, MAPkinase = mitogen-activated protein kinase, MHC = myosin heavy chain, mRNA = messenger RNA, NF-αB = nuclear factor-αB, SCI = spinal cord injury, SEM = standard error of the mean, TNF-α = tumor necrosis factor-α, VA = Department of Veterans Affairs, VL = vastus lateralis, VO2 = peak oxygen consumption.

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after stroke; (3) tumor necrosis factor-α (TNF-α) and inflammatory pathway activation as possible mediators of muscle atrophy and muscle injury; and (4) the beneficial effects of exercise interventions on skeletal muscle of individuals with stroke.

ALTERATIONS OF BODY COMPOSITION AND MUSCLE STRUCTURE AND FUNCTION IN AGING, INACTIVITY, AND AFTER SPINAL CORD INJURY

Adaptations in Skeletal Muscle Molecular Phenotype: Skeletal Muscle Plasticity

Mammalian skeletal muscle fibers have great adaptive potential. Muscle fibers have the ability to adjust their molecular, metabolic, and functional properties in response to altered functional demands, mechanical loading, or changes in neuromuscular activity. Myosin heavy chain (MHC) isoforms have different structural, enzymatic, and regulatory contractile properties and can thereby impart functional diversity to muscles. Slow-twitch muscle (MHC type I) isoform fibers are rich in mitochondria, resistant to fatigue, and less pH sensitive because of their highly oxidative metabolism [6–7]. Fast-twitch fibers vary in metabolic enzymes and fiber diameter. The larger fast-twitch fibers are glycolytic fibers (MHC type IIx), and the others are glycolytic/oxidative fibers (MHC type IIa). Both fast-twitch fiber types have larger diameters and faster force-generation capacity than slow-twitch fibers (MHC type I). The slow-twitch MHC type I isoform fibers are fatigue resistant, while the fast-twitch MHC type IIx isoform fibers fatigue easily and the fast-twitch MHC type IIa isoform fibers exhibit intermediate fatigue resistance. Regulation of MHC gene expression is controlled by a complex set of processes. Muscle fiber structural and functional characteristics are not fixed; they can be modified in response to several physiological and pathological conditions. Muscle MHC phenotype is regulated by growth hormone, insulin growth factor, thyroid, changes in load, innervation patterns, age-related changes, hypoxia, mitogen-activated protein kinase (MAP kinase) and stress pathway activation, electrical stimulation, and exercise [8–15]. The type and proportion of MHC isoforms expressed can serve as a cellular “marker” for muscle plasticity in response to perturbations. Muscle fiber properties can also be modified by pharmacological agents such as beta2-adrenergic agonists and corticosteroids. We would like to understand the factors regulating contractile protein, MHC, and skeletal muscle adaptations in response to altered use, loading states, and neural activation patterns after stroke. This information would inform efforts to assess the effect of various rehabilitation strategies that target skeletal muscle for chronic stroke.

Muscle Changes Related to Aging, Immobility, and Spinal Cord Injury

Sarcopenia of aging is a multifactorial process that affects skeletal muscle, including that of individuals with chronic stroke (Figure 1). Aging is associated with reduced strength, contraction velocity, and injury recovery [16–17]. Aging also results in decreased synthesis of myofibrillar components, increased production of catabolic cytokines, atrophy, and altered muscle metabolism [18–21]. With aging, motor unit dropout is coupled with increased motor unit size. The motor unit size increases because of reinnervation of adjacent denervated muscle fibers. Fast-twitch fibers are the most vulnerable to denervation and then reinnervation by slow-twitch motor neurons [22]. Normal aging leads to larger slow-twitch and smaller fast-twitch motor units, with an overall increase in slow-twitch fiber mass. This age-related increase in slow-twitch muscle is in opposition to our finding of increased fast-twitch MHC isoforms after stroke [23]. Aging also results in single fibers coexpressing multiple MHC isoforms [24]. Independent of age-related MHC isoform phenotype changes, calcium-activated myosin adenosine triphosphatase (ATPase) activity and maximum unloaded shortening velocity are reduced and contribute to age-related slowing of muscle contraction [25]. In humans, a decline in muscle strength begins at age 40 and is more dramatic after age 65. Although muscle mass decreases by 30 to 40 percent with sarcopenia of aging, muscle force and power decrease to a much greater extent, which suggests further alterations in muscle contractile properties and metabolism [20].

The disability of stroke leads to a relative inactivity, especially in the hemiparetic contralateral limb. Immobilized muscle has reduced eccentric, concentric, and isometric strength [26]. Physical inactivity results in reduced muscle mass and function, which parallel the declines that occur with aging. Muscle unloading produces a net deficit in quadriceps muscle total RNA, total messenger RNA (mRNA), and specific MHC mRNA levels, which can be partially restored with exercise [27].
Muscle immobilization reduces muscle fiber cross-sectional area [26] and downregulates MHC type I mRNA expression, while it upregulates MHC type IIx mRNA expression [26]. Physical inactivity produces similar degrees of muscle atrophy in young and old animals; however, recovery from the muscle atrophy is significantly delayed and reduced in older versus younger animals, thereby limiting recovery potential [16–17,28]. Exercise may prevent or delay sarcopenia by reducing the loss of muscle mass and strength through changes in myosin expression, even in aged mammals. However, the response to exercise is decreased in aged muscle. Because recovery from disuse atrophy is delayed with aging, minimizing the period of unweighting and immobilization is optimal. These biological processes that occur in muscle with aging and inactivity are essentially unexplored areas in stroke rehabilitation and have potentially great clinical relevance.

Skeletal muscle following stroke may be affected by the altered central neural activation and spasticity, similar to the muscle changes that occur after SCI. The weakness and spasticity have an interesting influence on the muscle, causing both reduced motor unit recruitment and excessive cocontraction, with an overactive stretch reflex. SCI induces a rapid loss of muscle mass and replacement with intramuscular fat [29]. MHC transcriptional activity is reduced and a shift to a fast-twitch MHC isoform profile occurs, especially an increase in MHC type IIx isoforms [30–31]. Skeletal muscle sodium-potassium ATPase concentration is reduced in spastic muscle [32]. The increased shortening velocity and fatigability of the paralyzed quadriceps muscles after SCI and other conditions that result in spasticity are probably secondary to the shift to fast-twitch MHC isoforms, reduced sodium-potassium ATPase concentration, reduced capillary density, and reduced oxidative capacity [32–36]. In addition, gene expression of enzymes that affect protein degradation (calpain-1 and enzymes associated with polyubquitination) is increased [30], which could contribute to muscle wasting after SCI. These findings suggest that muscle atrophy after SCI is likely a multifactorial process that affects transcription, translation, and protein degradation.
ALTERATIONS OF MUSCLE STRUCTURE
AND FUNCTION AFTER STROKE

Muscular Atrophy and Phenotype Shift After Stroke: Relation to Fitness and Function

Traditionally, upper motor neuron injury, as occurs in stroke, is not believed to result in muscular atrophy. Few studies have examined muscle abnormalities after stroke and their relationship to fitness and function. We examined relationships between gait deficit severity, peak oxygen consumption (VO\(_2\)), and body composition using dual X-ray absorptiometry (DXA) in 60 chronic hemiparetic stroke patients and found that lean mass of the contralateral limb was lower than that of the ipsilateral limb (mean ± standard error of the mean [SEM] = 8.3 ± 1.6 kg vs 8.6 ± 1.7 kg, \(p < 0.001\)) [37]. Stepwise regression revealed that both contralateral thigh lean mass (\(r = 0.61\)) and walking speed (cumulative \(r = 0.79\)) were independent and robust predictors of reduced fitness (VO\(_2\)) and accounted for 62 percent of the observed variance (\(p < 0.01\)). In contrast, the National Institutes of Health Stroke Scale, the modified Ashworth spasticity scale, stroke onset latency, and percent body fat were unrelated to VO\(_2\). These DXA results are substantiated by bilateral midthigh computed tomography cross-sectional area measurements in 30 chronic stroke patients. The contralateral midthigh area (mean ± SEM = 86.1 ± 29.3 cm\(^2\)) had 20 percent lower muscle cross-sectional area (\(p < 0.001\)) than the ipsilateral midthigh area (mean ± SEM = 100.9 ± 27.9 cm\(^2\)) (Figure 2) [38]. The reduced thigh muscle mass predicted lower VO\(_2\) in these patients [37]. The contralateral thigh muscle also had 25 percent higher intramuscular area fat deposition (\(p < 0.001\)) (Figure 2), a finding strongly associated with insulin resistance and its complications [38–40]. The reduced muscle mass and increased intramuscular fat are similar to recent findings in individuals with incomplete SCI [29]. Although these results suggest that reduced muscle mass is fundamentally related to poor fitness and physical performance capacity after stroke, they do not establish causality and do not account for reduced central muscle activation.

Altered Muscle Phenotype in Stroke

Little is known about skeletal muscle phenotypic abnormalities after stroke. Mechanisms for skeletal muscle alterations are probably similar in stroke and incomplete SCI, with reduced neuromuscular activation and muscle unloading but retained neuromuscular connectivity. Basic pathological studies reveal variable results with altered fiber type proportions, including selective fast-twitch MHC fiber atrophy and specific loss of slow-twitch MHC fibers in hemiparetic limbs of stroke patients [36,41–44]. The most comprehensive study by Landin et al. revealed (1) a shift to greater fast-twitch fiber proportions in the contralateral leg vastus lateralis (VL) muscle based on ATPase staining and (2) a reliance on anaerobic metabolism with rapid lactate generation during isolated contralateral or hemiparetic limb exercise in contrast to the oxidative metabolism during isolated ipsilateral leg exercise [44]. These findings are concordant with the major shift to fast-twitch MHC that we found in our recent study using routine ATPase staining at pH 4.6 and MHC immunohistochemistry and gel electrophoresis of bilateral leg VL muscle biopsies (Figures 3 and 4). Bilateral VL biopsies from 13 untrained stroke patients showed a significantly elevated proportion of fast-twitch

![Figure 2](image-url)

Computed tomography (CT) images show atrophy of (a) paretic leg midthigh muscle area compared with (b) nonparetic thigh. Low-density CT lean-tissue imaging of same individual shows increased intramuscular fat content in (c) paretic compared with (d) nonparetic thigh after stroke.
MHC isoforms in the contralateral (mean ± SEM = 68% ± 14%, range 46%–88% of total MHC) versus ipsilateral leg (mean ± SEM = 50% ± 13%, range 32%–76%, \( p < 0.005 \)) [23]. Interestingly, this shift to fast-twitch MHC composition in the contralateral muscle parallels that seen in animals and humans after SCI [45–46]. This result suggests that neurological alterations may be partially responsible for the muscle phenotype shift. The shift to fast-twitch MHC is in contrast to the shift to slow-twitch MHC in aging, where fast-twitch fibers are lost through denervation and slow-twitch fiber density increases through reinnervation [22]. The shift to fast-twitch MHC after stroke in contralateral leg muscle would be expected to result in a more fatigable muscle fiber type that could be more insulin resistant. In the contralateral leg only, the proportion of fast-twitch MHC isoform is strongly and negatively correlated to self-selected walking speed (\( r = -0.78, p < 0.005 \)), which suggests that neurological gait deficit severity may be a major contributor to MHC isoform expression and account for as much as 61 percent of its variance. The findings suggest that shifts in muscle phenotype may be fundamentally related to neuromuscular function. In our cohort, the muscular atrophy and the shift to the fast-twitch MHC isoform in the contralateral leg were both strong predictors of gait deficit severity [23,37]. Unfortunately, the current studies cannot explain the direction of these relationships; i.e., whether the atrophy and the shift in MHC expression are caused by or are a result of the gait deficit.

**TNF-α AND INFLAMMATORY PATHWAY ACTIVATION AS POSSIBLE MEDIATORS OF MUSCLE ATROPHY AND ALTERED MUSCLE METABOLISM AFTER STROKE**

TNF-α and nuclear factor-κB (NF-κB) have been implicated with muscular atrophy in models of disuse, cachexia, and sarcopenia. TNF-α may contribute to atrophy through a number of mechanisms, including inhibition of protein synthesis and reduced expression of MyoD, a transcriptional regulator of myofiber gene expression and accelerated protein breakdown through activation of ubiquitin proteases and NF-κB and apoptotic cell death [47–52]. TNF-α appears to preferentially downregulate adult slow-twitch MHC protein synthesis and enhance its degradation [53], which may in part explain our findings of elevated fast-twitch MHC in the hemiparetic leg. TNF-α also activates NF-κB transcriptional factor [54], which may increase inducible nitric
associated with muscle atrophy [64]. These inflammatory proteins, and performance.

mediators can negatively affect muscle mass, structural proteins. Elevated TNF-α could mediate atrophy, accelerate oxidative injury, and alter expression that could potentially influence muscle structure and function [63]. The presence of reactive oxidative species disturbs muscle redox status and can result in muscle fatigue and/or injury and further activates NF-κB [62]. Cytokine-independent activation of NF-κB is also associated with muscle atrophy [64]. These inflammatory mediators can negatively affect muscle mass, structural proteins, and performance.

Although TNF-α is negligibly expressed in skeletal muscle, we found that TNF-α mRNA expression is elevated in the contralateral VL muscle of patients with stroke compared with the ipsilateral leg of patients with stroke and age-matched nonneurological control subjects [65]. TNF-α mRNA levels were three times higher in contralateral leg muscle of patients with stroke than in control muscles. A trend existed for almost two times higher TNF-α in the ipsilateral leg muscle compared with nondisabled control subjects. The finding of elevated TNF-α in both the contralateral and ipsilateral leg muscles suggests a systemic as well as local inflammation that could augment hemiparetic leg muscular atrophy and increase insulin resistance after stroke. Three-quarters of these subjects with stroke and elevated skeletal muscle TNF-α were on anti-inflammatory medications, such as aspirin, or hydroxy-3-methylglutaryl-CoA reductase medications and insulin sensitizers, which suggests that these medications are not capable of completely countering inflammation at the level of the skeletal muscle. We also found, using a complementary DNA NF-κB signaling gene miniarray (GE SuperArray, General Electric, Co; Fairfield, Connecticut), that NF-κB inflammatory pathway gene activation is differentially upregulated in the contralateral compared with the ipsilateral VL muscle (N = 6). Products of NF-κB activation could mediate atrophy, accelerate oxidative injury, and alter important structural proteins. Elevated TNF-α protein and mRNA in frail elderly skeletal muscles can be decreased with strength training [66]. Thus, interventions aimed at attenuating elevated skeletal muscle inflammatory pathways may represent new targets for reducing both disability and cardiovascular disease risk after stroke.

A remarkably large incidence of insulin resistance and type 2 diabetes is present in individuals who had stroke [67–68]. New evidence from the Dutch Transient Ischemic Attack Trial reveals a two- and threefold increased risk of stroke for individuals with impaired glucose tolerance and type 2 diabetes, respectively [68]. Circulating levels of TNF-α and interleukin-6 are elevated in subjects with type 2 diabetes and impaired glucose tolerance [69–70]. Systemic and muscle-specific elevations in TNF-α are strongly linked to insulin resistance and diabetes [71]. TNF-α directly impairs insulin signaling in muscle [72] and is inversely related to maximum glucose disposal rate [71–74]. Exercise can reduce TNF-α and improve skeletal muscle metabolism and systemic insulin sensitivity [75]. Hence, better understanding of the molecular mechanisms of accelerated inflammation in the contralateral leg muscle of subjects with stroke and effects of exercise may have important implications for cardiovascular health of people who remain at high risk for stroke recurrence.

**BENEFICIAL EFFECTS OF EXERCISE INTERVENTIONS ON SKELETAL MUSCLE IN CHRONIC STROKE**

A number of aerobic and resistive exercise training strategies have proven beneficial for patients with chronic stroke. These exercise strategies include treadmill, robot-assisted walking, and strength training [76–80]. Exercise may have many beneficial effects at the level of skeletal muscle after stroke. It may prevent changes associated with physical inactivity. Exercise in nonneurological populations and animal models can prevent or delay sarcopenia through changes in myosin expression and reduce loss of muscle mass and strength [37,81]. Elevated TNF-α protein and mRNA in frail elderly skeletal muscles can be decreased with strength training [66]. Aerobic exercise can produce skeletal muscle adaptations that protect myocytes and muscle fibers from muscle injury, improve muscle performance, and delay muscle fatigue [27,81–84]. To our knowledge, the effects of exercise on muscle structure and function have never been systematically studied after stroke.

We and others have reported that treadmill aerobic exercise, as a task-oriented training model, improves fitness and mobility function in patients with chronic stroke [76,85–90]. Our results show that treadmill exercise is superior to a program with components of “conventional rehabilitation therapy” for improving fitness and mobility.
function levels in patients with chronic stroke [85–90].
Aerobic exercise can induce profound molecular changes in “neurologically intact” muscle [91], promoting fast- to slow-twitch MHC fiber conversion. Treadmill exercise had a significant effect on MHC expression. The total MHC concentration and proportion of slow- and fast-twitch MHC type Ila isoforms in the contralateral limb significantly increase after 6 months of treadmill exercise (Figure 5).* In contrast, the control stroke group that was enrolled in a 6-month time-matched stretching program did not have a significant change in the MHC concentration or profile distribution. In a small cohort of patients with chronic stroke, these data suggest that treadmill exercise can reverse the contralateral leg MHC profile abnormalities. The beneficial effects of aerobic exercise training have also been demonstrated in SCI [83,92–93]. Functional electric stimulation can induce skeletal muscle changes, such as a shift to a fast-twitch MHC phenotype in SCI [94–95]. Transcranial electrical nerve stimulation can improve H reflex response and reduce spasticity after stroke and, in combination with task-related training, improve walking function [46,96]. Finally, pharmaceutical agents can also target skeletal muscle. Beta2-adrenergic agonists can induce skeletal muscle hypertrophy by inducing myocyte proliferation, myogenic differentiation, satellite cell recruitment into muscle fibers, and the initiation of translation that increases protein synthesis [97–98]. As our understanding of the ability of exercise rehabilitation strategies to improve skeletal muscle structure and function increases, we will be better poised to prescribe specific rehabilitation protocols that reduce the musculoskeletal component of disability after stroke.

**SUMMARY**

Stroke is the leading cause of disability in the United States. This disability is traditionally attributed to the brain injury and the high risk for recurrent stroke ascribed to preexisting cardiovascular disease risk factors. We propose a new hypothesis that secondary biological abnormalities of muscle atrophy, altered contractile protein expression, and inflammation in the contralateral leg skeletal muscle propagate disability. While animal models and clinical data show that both disuse and abnormal neural innervation can produce such abnormalities, little is known about their presence after stroke and their potential for reversal with exercise. We report altered major structural skeletal muscle proteins and activation of inflammatory pathways in bilateral VL muscle biopsies from untrained hemiparetic stroke patients. Contralateral leg muscle biopsies obtained after the 6-month treadmill exercise program can help us determine the training components of the aerobic treadmill exercise program that produce the greatest beneficial adaptations in skeletal muscle. Study results may provide a biological rationale and new molecular targets for novel pharmacological therapeutics aimed at improving muscle structure and metabolic function after stroke. The biological relevance and potential for clinical applications in both disability reduction and secondary cardiovascular disease prevention will likely extend to other neurological conditions that engender disuse and abnormal neural innervation, such as multiple sclerosis, closed head injury, and SCI.

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